

Transformation of Cholanic Acid Derivatives into Pharmacologically Active Esters of Phenolic Acids by Heterogeneous Wittig Reaction

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Steroid esters of cinnamic acid derivatives have been synthesized by a heterogeneous Wittig reaction under sonochemical conditions from the corresponding triphenylphosphonium bromides and unprotected phenolic aldehyds using K_2CO_3 as a base. 5β -Cholan-3 α , 7 α , 12 α , 24-*E*-ferulate (**11'**) exhibited a marked inhibitory effect on influenza virus A. The synthetic 3 α , 24-*E*-diferulates of 5β -cholan-3 α , 24- diol, 5β -cholan-3 α , 12 α , 24-triol and 5β -cholan-3 α , 7 α , 12 α , 24-tetrol (**8**, **9** and **12**) showed antitumor activity on leukemia P-388 in mice.

Introduction

In the course of the continuing search for biologically active agents from plants many new triterpene monoesters of phenylpropanoic acids were found (Tommasi *et al.*, 1992; Pan *et al.*, 1994). Recently the isolation and characterization of two new cytotoxic di-*p*-coumaroyl triterpenes from *Ilex asprella* was reported (Kashiwada *et al.*, 1993). The study of the general pharmacological activities of these natural compounds has been seriously hampered by their limited amount available from natural sources.

Recently in our laboratory a new synthesis of cholesteryl esters of cinnamic acids derivatives was elaborated, using the Wittig reaction in solid-liquid heterogeneous medium under sonochemical conditions (Elenkov *et al.*, 1995). The results of work prompted us to develop this method for preparation of compounds with more than one ester group starting from polyhydroxylated steroids and to test their biological activity.

This paper describes our work on the sonochemical Wittig reaction between different phenolic aldehydes and phosphonium salts of the corresponding halides of 5β -cholanol derivatives as well as

the antiviral and antitumor study on the synthesized compounds.

Experimental

¹H-NMR spectra were recorded on Bruker 250 MHz for solutions in $CDCl_3$ or CD_3OD with TMS as internal standard. The UV spectra in EtOH solutions were measured with a Specord UV-VIS spectrophotometer.

General procedure for preparation of 5β -cholanols

14.2 mmol of Me ester of cholanic acid (cholic, deoxycholic, lithocholic) were dissolved in 100 ml dry THF and added under argon to 42 mmol $LiAlH_4$ in 20 ml THF. The reaction mixture was stirred 10 h at room temperature. At the end of reaction 20 ml 10% NH_4Cl were added. The residue was filtered and washed with EtOAc. The organic phase was washed with water, dried over Na_2SO_4 , evaporated and subjected to CC or recrystallization from acetone.

5β -cholan-3 α ,24-diol (**1**), 80% yield, m.p. 177–176 °C

¹H-NMR ($CDCl_3$) 0.65 (3H, s, Me-18), 0.91 (3H, s, Me-19), 0.93 (3H, d, $J = 6.4$, Me-21), 3.61 (3H, m, H-3 β , 24- CH_2 -O).

5β -cholan-3 α , 12 α , 24-triol (**2**). 80% yield, m. p. 199–201 °C

¹H-NMR ($CDCl_3$) 0.68 (3H, s, Me-18), 0.91 (3H, s, Me-19), 0.99 (3H, d, $J = 6.4$, Me-21), 3.61 (3H, m, H-3 β , - CH_2 -OH-24), 3.99 (1H, m, H-7 β)

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5 β -cholan-3 α , 7 α , 12 α , 24-tetrol (**3**). 89% yield, m.p. 206–208 °C

¹H-NMR (CDCl₃ : CD₃OD = 1:1) 0.71 (3H, s, Me-18) 0.98 (3H, s, Me-19), 1.015 (3H, d, *J* = 6.6, Me-21), 3.45 (1H, m, H-3 β), 3.54 (2H, t, *J* = 6.8, 24-CH₂-), 3.83 (1H, bs, H-7 β), 3.98 (1H, bs, H-12 β).

General procedure for esterification of 5 β -cholanols with BrCH₂COOH

0.28 mmol 5 β -cholanol, 0.75 mmol bromoacetic acid, 0.75 mmol DCC and 0.0112 mmol 4-DMAP were dissolved in 20 ml dry THF. The reaction mixture was stirred under argon at room temperature for 1 h. The residue of dicyclo hexylurea was filtered and washed with CHCl₃ and the combined solutions evaporated under vacuum. The residue was purified by CC on silica.

5 β -cholan-3 α ,24-diol-3 α ,24-dibromoacetate (**4**), oil, 85% yield

¹H-NMR (CDCl₃) 0.65 (3H, s, Me-18), 0.91 (3H, s, Me-19), 0.93 (3H, d, *J* = 6.2, Me-21), 3.80 (2H, s, -OCOCH₂Br), 3.82 (2H, s, -OCOCH₂Br), 4.15 (2H, t, -CH₂-24), 4.75 (1H, m, H-3 β).

5 β -cholan-3 α ,12 α ,24-triol-3 α ,24-dibromoacetate (**5**), oil, 50% yield

¹H-NMR (CDCl₃) 0.69 (3H, s, Me-18), 0.93 (3H, s, Me-19), 0.99 (3H, d, *J* = 6.4, Me-21), 3.79 (2H, s, -OCOCH₂Br), 3.83 (2H, s, -OCOCH₂Br), 4.00 (1H, bs, H-12 α), 4.15 (2H, t, -CH₂-24), 4.78 (1H, m, H-3 β).

5 β -cholan-3 α ,7 α ,12 α ,24-tetrol-24-monobromoacetate (**6**), oil, 22% yield

¹H-NMR (CDCl₃:CD₃OD 1:1) 0.71 (3H, s, Me-18), 0.91 (3H, s, Me-19), 3.43 (1H, m, H-3 β), 3.83 (1H, bs, H-7 β), 3.88 (2H, s, -OCOCH₂Br), 3.97 (1H, bs, H-12 β), 4.16 (2H, t, *J* = 6.2, -CH₂-24).

5 β -cholan-3 α ,7 α ,12 α ,24-tetrol-3 α ,24-dibromoacetate (**7**), oil, 55% yield

¹H-NMR (CDCl₃:CD₃OD 1:1) 0.71 (3H, s, Me-18), 0.92 (3H, s, Me-19), 1.00 (3H, d, *J* = 6.4, Me-21), 3.79 (2H, s, OCOCH₂Br), 3.83 (2H, s, -OCOCH₂Br), 3.86 (1H, bs, H-7 β), 4.01 (1H, bs, H-12 β), 4.13 (2H, t, *J* = 6.7, -CH₂-24), 4.65 (1H, m, H-3 β).

Preparation of phosphonium salts

0.25 mmol dibromide and 0.56 mmol triphenylphosphine were dissolved in 30 ml dry THF. After

72 h at room temperature the residue was filtered, dissolved in MeOH and extracted twice with *n*-hexane. The lower layer was evaporated to dryness. Yields: 92%.

General procedure for synthesis of diesters

A solution of 0.234 mmol of phosphonium salt in 0.5 ml CHCl₃ and a solution of aromatic aldehyde in 0.5 ml 1,4-dioxane were mixed and added to 0.35 mmol of K₂CO₃. The reaction mixture was sonicated in a Lechpan Type UM 0.5 ultrasonic bath at 25 °C. The reaction was monitored by TLC (Alufolien Kieselgel 60F₂₅₄, Merck, *n*-hexane/acetone). The reaction mixture was washed successively with 5% HCl and H₂O. The organic phase was dried over Na₂SO₄, evaporated to dryness and subjected to CC (silica). All products were characterized by their UV- and ¹H-NMR spectra.

5 β -cholan-3,24-diol-3 α ,24-di-*E*-ferulate (**8**)

UV (MeOH) λ_{\max} 233.5, 300, 314.2 nm.

¹H-NMR (CDCl₃) 0.66 (3H, s, Me-18), 0.94 (3H, s, Me-19), 3.90 (3H, s, -OMe), 3.92 (3H, s, -OMe), 4.14 (2H, m, CH₂-24), 4.80 (1H, m, H-3 β), 5.95 (1H, bs, Ar-OH), 6.26 (1H, d, *J* = 15.9, H-2'), 6.28 (1H, d, *J* = 15.9, H-2'), 6.91 (2H, m, H-6''), 7.05 (4H, m, H-2'', H-3''), 7.56 (1H, d, *J* = 15.9, H-3'), 7.60 (1H, d, *J* = 15.9, H-3').

5 β -cholan-3 α ,12 α ,24-triol-3 α ,24-di-*E*-ferulate (**9**)

UV (MeOH) λ_{\max} 234.8, 300, 326.2 nm.

¹H-NMR (CDCl₃) 0.70 (3H, s, Me-18), 0.94 (3H, s, Me-19), 1.01 (3H, d, *J* = 6.3, Me-21), 3.92 (3H, s, -OMe), 3.93 (3H, s, -OMe), 4.02 (1H, bs, H-12 β), 4.17 (2H, m, -CH₂-24), 4.8 (1H, m, H-3 β), 5.86 (1H, s, Ar-OH), 6.25 (1H, d, *J* = 15.9, H-2'), 6.28 (1H, d, *J* = 15.9, H-2'), 6.90 (2H, m, H-6''), 7.14 (4H, m, H-2'', H-3''), 7.58 (1H, d, *J* = 15.9, H-3'), 7.60 (1H, d, *J* = 15.9, H-3').

5 β -cholan-3 α ,7 α ,12 α ,24-tetrol-3 α ,24-di-*E*-cinnamate (**10**)

UV (MeOH) λ_{\max} 280.

¹H-NMR (CDCl₃) 0.72 (3H, s, Me-18), 0.93 (3H, s, Me-19), 3.87 (1H, bs, H-7 β), 4.03 (1H, bs, H-12 β), 4.2 (2H, m, -CH₂-24), 4.72 (1H, m, H-3 β), 6.39 (1H, d, *J* = 16.0, H-2'), 6.44 (1H, d, *J* = 16.0, H-2'), 7.38 (5H, m), 7.53 (5H, m), 7.65 (1H, d, *J* = 16.0, H-3'), 7.68 (1H, d, *J* = 16.0, H-3').

5 β -cholan-3 α ,7 α ,12 α ,24-tetrol-3 α ,24-di-*E*-ferulate (**11**), oil

UV (EtOH) λ_{\max} 227.5, 290, 342 nm.

$^1\text{H-NMR}$ (CDCl_3) 0.73 (3H, s, Me-18), 0.96 (3H, s, Me-19), 1.05 (3H, d, $J = 6.3$, Me-21), 3.82 (1H, bs, H-7 β), 3.88 (6H, s, -OMe), 3.98 (1H, bs, H-12 β), 4.16 (2H, t, -CH₂-24), 4.66 (1H, m, H-3 β), 6.50 (1H, d, $J = 16.0$, H-2'), 6.55 (1H, d, $J = 16.0$, H-2'), 6.77–6.83 (2H, m), 6.93–6.94 (2H, m), 6.97–7.20 (2H, m), 7.95 (1H, d, $J = 16.0$, H-3'), 7.98 (1H, d, $J = 16.0$, H-3').

5 β -cholan-3 α ,7 α ,12 α ,24-tetrol-24-*E*-ferulate (**11'**), oil

UV (EtOH) λ_{\max} 227.5, 287.5, 333.3 nm.

$^1\text{H-NMR}$ (CDCl_3 : CD_3OD 1:1) 0.70 (3H, s, Me-18), 0.90 (3H, s, Me-19), 1.01 (3H, d, $J = 6.5$, Me-21), 3.46 (1H, m, 1H-3 β), 3.85 (1H, bs, H-7 β), 3.92 (3H, s, -OMe), 4.00 (1H, s, H-12 β), 4.16 (2H, m, -CH₂-24), 6.71 (1H, d, $J = 16.2$, H-2'), 6.86 (2H, m, H-5'', H-6''), 7.10 (1H, m, H-2''), 7.94 (1H, d, $J = 16.2$, H-3').

5 β -cholan-3 α ,7 α ,12 α ,24-tetrol-3 α ,24-di-*E*-ferulate (**12**), oil

UV (MeOH) λ_{\max} 235.2, 299, 325 nm.

$^1\text{H-NMR}$ (CDCl_3) 0.72 (3H, s, Me-18), 0.93 (3H, s, Me-19), 1.024 (3H, d, $J = 6.4$, Me-21), 3.87 (1H, bs, H-7 β), 3.92 (3H, s, -OMe), 3.93 (3H, s, -OMe), 4.02 (1H, bs, H-12 β), 4.18 (2H, m, -CH₂-24), 4.73 (1H, m, H-3 β), 5.88 (1H, s, Ar-OH), 6.24 (1H, d, $J = 15.9$, H-2'), 6.28 (1H, d, $J = 15.9$, H-2'), 6.91 (1H, m, H-6''), 7.06 (2H, m, H-2'', H-3''), 7.56 (1H, d, $J = 15.9$, H-3'), 7.63 (1H, d, $J = 15.9$, H-3').

Bioassays

Viruses

Poliovirus (Mahoney strain) (PV1) (WHO Regional Reference Laboratory, Institute of Poliomyelitis and Viral Encephalitis, Vnukovo, Moscow District, Russia), influenza virus A/chicken/Germany/27/Weybridge (H7N7) (FPV) (Institute of Virology, Bratislava, Slovak Republic), Newcastle disease virus (Russeff strain) (NDV), (Central Veterinary Research Institute, Sofia) and pseudorabies virus (PsRV, Aujeszky, A2 strain) (Central Veterinary Research Institute, Sofia) were used.

Cell cultures

FL cells were grown in a medium containing 10% heated calf serum in a mixture of equal parts of medium 199 (Difco) and Hanks's saline, supple-

mented with penicillin (100 IU/ml) and streptomycin (100 $\mu\text{g/ml}$).

Primary chick embryo fibroblasts cultures (CEC) were prepared after Porterfield (1960) and the cell suspension, $1-1.5 \times 10^6$ cells/ml, was seeded in a growth medium Eagle's MEM (Difco) supplemented with 10% calf serum.

Antiviral tests

The agar-diffusion plaque-inhibition method with cylinders (Rada and Zavada, 1962) was used for the initial screening for antiviral activity and was performed as described previously (Galabov *et al.*, 1996). The compounds tested (0.1 ml of 0.5% w/v solutions in DMSO) were added dropwise in the 6-mm glass cylinders fixed in the agar overlay. The antiviral effect (E) was recorded on the basis of the difference between the size (diameter (ϕ), in mm) of the zone of plaque inhibition (ϕ_i) and the zone of cytotoxicity (ϕ_c) and designated as follows: -, $\Delta\phi = 5$ mm; +, $\Delta\phi = 6-10$ mm; ++, $\Delta\phi = 11-20$ mm; +++, $\Delta\phi = 21-40$ mm; +++++, $\Delta\phi > 40$ mm.

The compounds showing a marked antiviral effect ($\Delta\phi > 10$ mm) were then studied in the cytopathic effect (CPE) inhibition multicycle test on monolayer cell cultures in 96-well plastic microplates (Flow) following the setup described by Galabov *et al.* (1996), the minimal 50% inhibitory concentration (MIC_{50}) value being determined.

Cytotoxicity test

The compound tested was added to the growth medium (10% calf serum and antibiotics in Eagle's MEM (Flow) for CEC) immediately before cell seeding in 24-well plastic microplates (three wells per sample). The compound cytotoxicity was assessed by following the course of cell growth until the stationary phase, when the cell growth 50% inhibitory concentration (CGIC_{50}) was evaluated.

Antitumor test

Tumor passages and therapeutic experiments in vivo were carried out according to the standard methods (Sofina *et al.*, 1980). Leukemia P-388 was used as transplantable tumor model. Leukemia P-388 was propagated in a peritoneal cavity of male DBA₂ mice with 10^6 cells/mouse on day 0. For the experiments transplanted BDF₁ mice of both sexes were treated i.p. with different doses of compounds to determine the maximum active one.

The animals were divided in experimental groups of six mice each and a control one. The

treatment was performed on the 24th hour after transplantation. The mice had been treated for 5 successive days. The samples were dissolved in saline to which 1–2 drops of Tween-80 were added to increase their solubility. The biological activity of the compounds was assessed by the increasing life span (ILS) of the mice at 125% minimum criteria for activity.

Results and Discussion

5 β -Cholanols obtained by LiAlH₄ reduction of methyl esters of easily available cholic acids (cholic, deoxycholic and lithocholic) were used as starting steroids. Since the reactivity order of the hydroxyl groups of the obtained 5 β -cholanols (Fig. 1) is 24 > 3 α > 7 α > 12 α , the conditions have to be found to direct the acylation of **2**

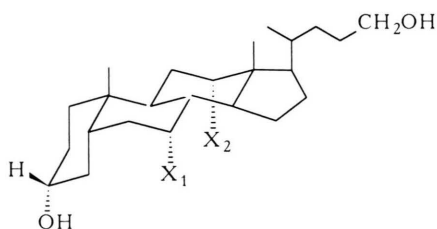
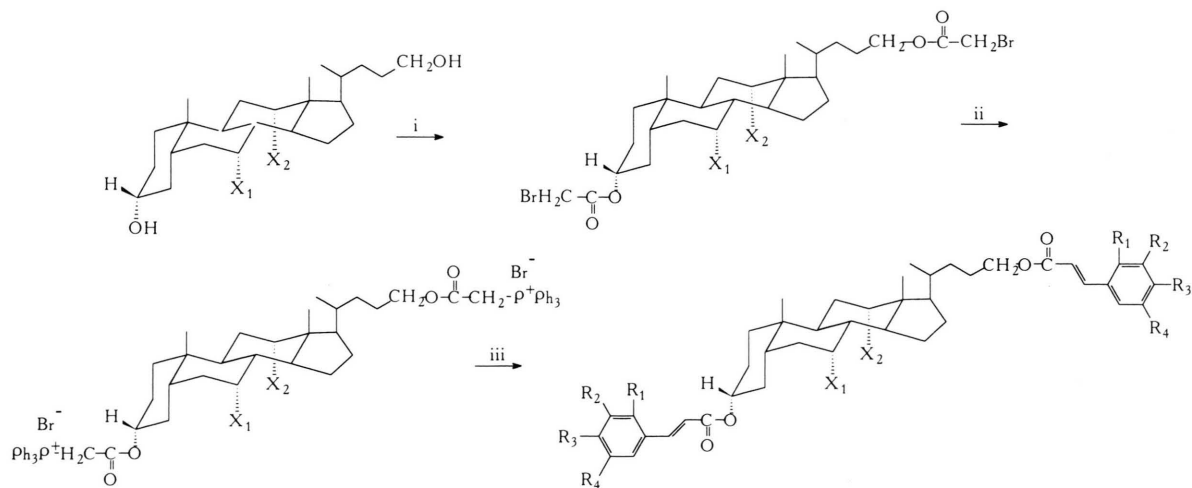


Fig. 1.
1. X₁ = X₂ = H
2. X₁ = H, X₂ = OH
3. X₁ = X₂ = OH

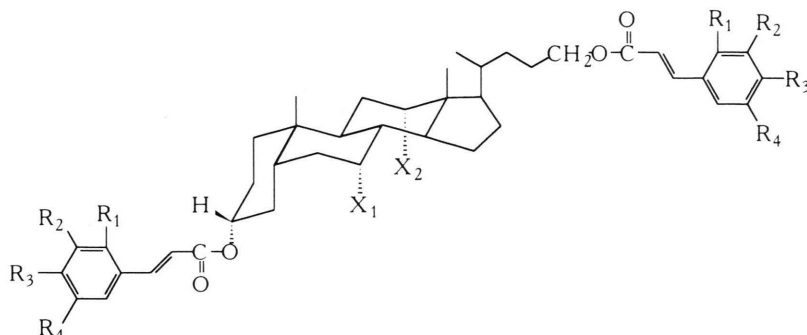
and **3** to an optimal quantity of the desired 5 β -cholan-3 α ,12 α , 24-triol-3 α ,24-dibromoacetate and 5 β -cholan-3 α ,7 α , 12 α ,24-tetrol-3 α ,24-dibromoacetate (Scheme 1). After much experimentation it was found that acylation of 5 β -cholan-3 α ,7 α ,12 α ,24-tetrol gave the desired 3 α ,24-dibromoacetate in 55% yield (22% 24-monobromoacetate and 13% tribromoacetate) by optimal ratio of tetrol: BrCH₂COOH:DCC:4-DMAP = 0.16:0.42:0.42:0.006 and reaction time 1h. By running of the reaction in an absence of 4-DMAP from the reaction mixture were isolated the starting tetrol, significant amount of 24-monobromoacetate and traces 3 α ,24-dibromoacetate.

The Wittig reaction of the phosphonium salts of the corresponding dibromides and different phenolic aldehydes (Scheme 1) was carried out under sonochemical condition in heterogeneous solid-liquid medium with K₂CO₃ as a base. The results obtained are shown in Table I. In the ¹H-NMR spectra of the synthesized diesters two one proton doublets at $\delta \approx 6.25$ –6.55 with $J \approx 16$ Hz and two another one proton doublets with the same J const. at $\delta \approx 7.56$ –7.98 were observed, characteristic for two -CH=CH- *trans* groups. The content of the *Z*-isomer in the products obtained is less than 5%, as estimated by ¹H-NMR spectroscopy. A monoester at C-24 of 5 β -cholan-3 α ,7 α ,12 α ,24-tetrol with 2-hydroxy-3-methoxycinnamic acid **11'** was synthesized under the same conditions.



Scheme 1. i, tetrahydrofuran, N,N - dicyclohexylcarbodiimide, 4 - dimethylaminopyridine; ii, tetrahydrofuran, triphenylphosphine; iii, aromatic aldehyde, chloroform - 1,4 - dioxane (1:1), K₂CO₃, sonication.

Table I. Synthesis of steroid diesters of submitted cinnamic acid under sonochemical conditions.



Compound	X ₁	X ₂	R ₁	R ₂	R ₃	R ₄	Yields, %	Reaction time [h]
8	H	H	H	OCH ₃	OH	H	45	12
9	H	OH	H	OCH ₃	OH	H	38	12
10	OH	OH	H	H	H	H	50	4
11	OH	OH	OH	OCH ₃	H	H	91	24
12	OH	OH	H	OCH ₃	OH	H	51	12

Table II. Screening of synthetic esters for antiviral activity by agar-diffusion plaque-inhibition test^a.

Compd.	Dose [mm] ^b	PV1			FPV			NDV			PsRV		
		φ _i	φ _t	E	φ _i	φ _t	E	φ _i	φ _t	E	φ _i	φ _t	E
11'	8.8	0	6.8	-	22.7	11.2	+	20.2	10.4	+	25.0	17.5	+
11	6.7	0	9.2	-	18.7	10.0	±	18.0	10.1	+	21.9	15.5	+
12	6.7	0	7.5	-	0	7.0	-	0	8.5	-	10.2	9.7	-
9	6.8	0	6.7	-	0	8.0	-	0	7.7	-	0	9.2	-
10	7.6	0	7.3	-	0	7.7	-	0	8.0	-	0	9.2	-

^a Φ_i = diameter of inhibition zone (mm); Φ_t = diameter of toxicity zone (mm); E = antiviral effect.

^b Per cylinder (dissolved in 0.1 ml DMSO).

PV1, poliovirus; FPV, influenza virus; NDV, Newcastle disease virus; PsV, pseudorabies virus.

The method reported in this paper proceed without protection of the phenolic groups of the aromatic aldehydes. In most cases the yields were 40%- 91% with reaction time of 4–24 h.

Antiviral activity

The compounds **9**, **10**, **11**, **11'** and **12** were tested for antiviral activity in vitro against representatives of four taxonomic viral groups, namely, picorna-, orthomyxo-, paramyxo- and herpes viruses. The viruses used were PV1, FPV, NDV and PsRV, respectively.

As seen in Table II, compound **11'** demonstrated a marked activity against FPV in the agar-diffusion plaque-inhibition test. This effect was

well reproducible also in the CPE inhibition test. MIC₅₀ value of 3.2 μg/ml was determined and selectivity ratio (CGIC₅₀/MIC₅₀) of 10.9 been evaluated. Compound **11** showed a borderline effect towards FPV and both compounds showed limited effects against NDV and PsRV in the screening test. The rest of the compounds was not active.

Antitumor activity

The synthetic esters **8**, **9** and **12** were investigated for antitumor activity on leukemia P-388 in mice. The inhibitory action of the compounds was evaluated by percentage of ILS (increasing life span) of the animals. Criteria for antitumor activity of the substances is ≥125%. The results were

Table III. Antitumor action of synthetic esters on leukemia P-388.

Compound	Dose [mg/kg]	MST[days]* Control	ILS** %
12	2.0	11.8/10.5	112.4
	3.0	10.3/9.0	114.0
	4.0	11.8/10.5	115.1
	6.0	10.8/9.0	120.0
	9.0	11.8/9.0	131.0
	10.0	11.4/9.3	122.6
	12.0	10.9/9.0	121.0
	14.0	10.6/9.2	116.0
8	2.0	11.0/10.5	104.8
	4.0	9.5/9.0	105.5
	5.0	11.2/10.5	106.6
	6.0	10.6/9.0	117.7
	8.0	11.5/9.0	127.8
	10.0	10.2/9.0	113.4
	12.0	10.3/9.2	111.9
	14.0	10.2/9.3	110.0
9	2.0	11.8/10.5	112.4
	4.0	10.5/9.0	116.6
	5.0	16.4/10.5	156.2
	6.0	11.3/9.2	122.8
	8.0	10.8/9.0	120.0
	10.0	10.2/9.0	113.3

* MST – mean survival time.

** ILS – increased life span.

shown in Table III. As seen in the table all three compounds possessed antitumor activity as follow: **12** at a dose of 9 mg/kg ILS of the mice 130.33%; **8** at a dose of 8 mg/kg ILS – 127.77% and **9** at a dose of 5 mg/kg ILS – 156.19%.

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Elenkov I., Todorova D., Bankova V. and Milkova Ts. (1995), Synthesis of steryl esters of phenolic acids by a heterogeneous Wittig reaction. *J. Nat. Prod.* **58**, 280–283.

Kashiwada Y., Zhang De-Cheng, Chen Yun-Pan, Cheng Chang-Ming, Chen Hsien-Tsung, Chang Hsien Chang, Chang Jer-Jang and Lee Kuo-Hsuing. (1993), Antitumor agents, 145. Cytotoxic asprellic acids A and C and asprellic acid B, new *p*-coumaroyl triterpenes from *Ilex asprella*. *J. Nat. Prod.* **56**, 2077–2082.

De Tommasi N., De Simone Fr., Pizza C., Mahmood N., Moore P., Conti C., Orsi N. and Stein M. (1992). Constituents of *Eriobotrya japonica* a study of their antiviral properties. *J. Nat. Prod.* **55**, 1067–1073.

Pan H., Lundgren L. and Anderson R. (1994), Triterpene caffeates from bark of *Betula pubescens*. *Phytochemistry* **37**, 795–799.

Rada B. and Zavada J. (1962), Screening test for cytostatic and virostatic substances. *Neoplasma* **9**, 57.

Galabov A. S., Iossifova T., Vassileva E. and Kostova I. (1996), Antiviral activity of some hydroxycoumarin derivatives. *Z. Naturforsch.* **51c**, 558–562.

Sofina Z. and Sirkin A. (1980), Experimental evaluation of antitumor drugs in USSR and USA. Moscow, Medicina, 296p.